

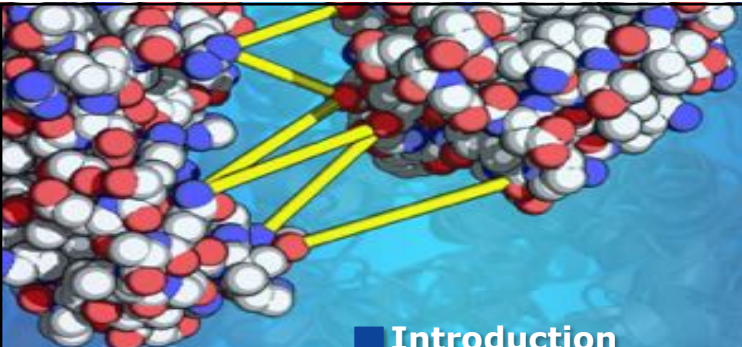
Unravelling the social life of proteins by integrative modelling

Prof. Alexandre M.J.J. Bonvin
Bijvoet Centre for Biomolecular Research
Faculty of Science, Utrecht University
the Netherlands
a.m.j.j.bonvin@uu.nl
@amjjbonvin

Utrecht Bioinformatics Center

CROP INNOVATION & BUSINESS
April 12 - 13, 2021 | Online Edition

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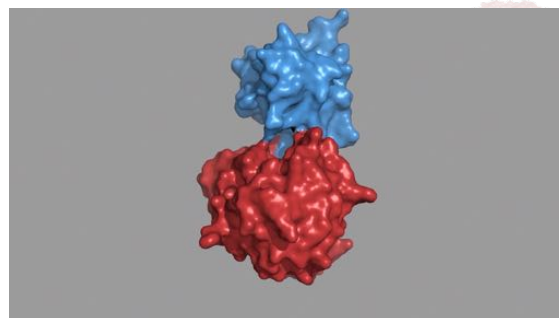


Overview

- Introduction
- Information-driven docking with HADDOCK
- A few examples
- Potential and perspectives in crop research

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Understanding life at the molecular level



Applications:

- Disease-causing mutations
- Engineer better molecules for material, health or food applications
- Obtain a starting point for drug design to combat disease

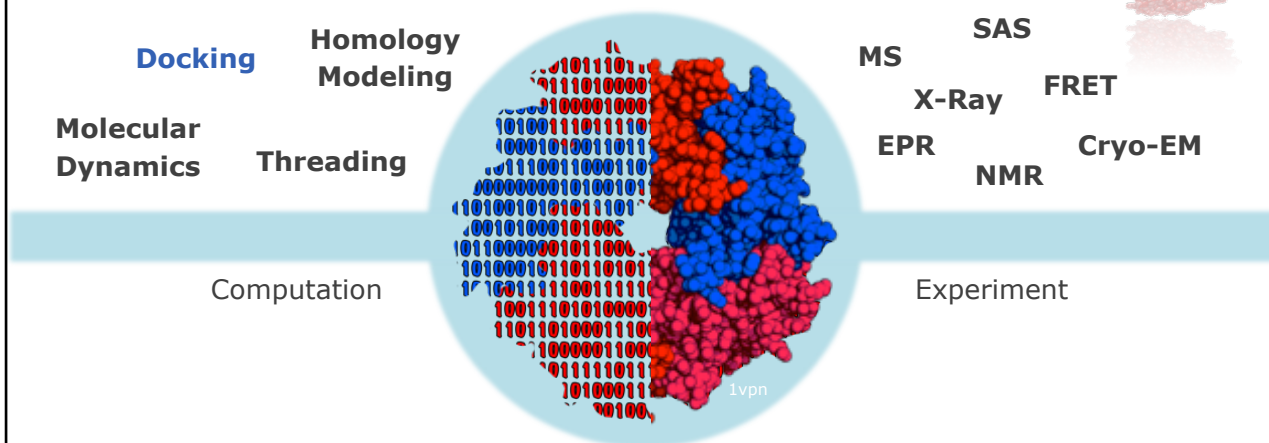


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Structural biology of interactions

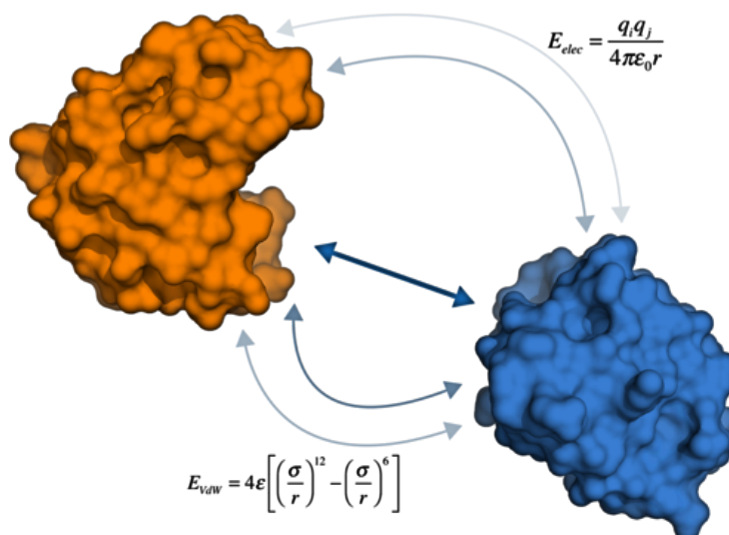


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Solving 3D puzzles by molecular docking



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HADDOCK: An integrative modeling platform

Incorporates ambiguous and low-resolution data to aid the docking

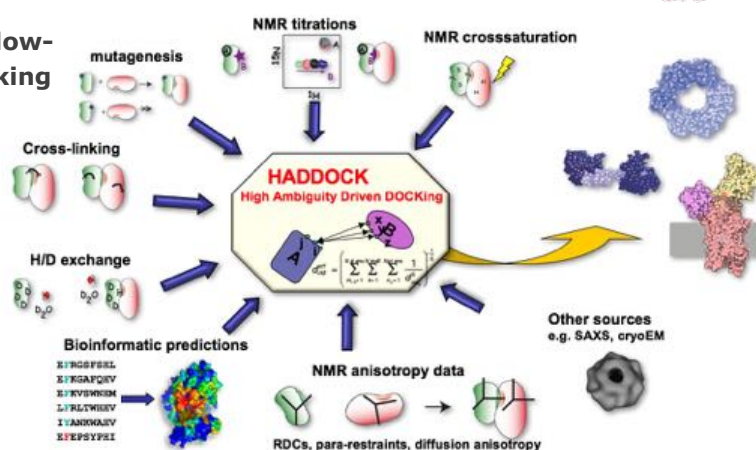
Capable of docking up to 20 molecules (new version)

Symmetries can be leveraged

Allows for flexibility at the interface

Final flexible refinement in explicit solvent

One of the best performing software in CAPRI



Dominguez, Boelens & Bonvin. JACS 125, 173 (2003).




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<http://www.bonvinlab.org/software>


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HADDOCK


it0


it1



itw

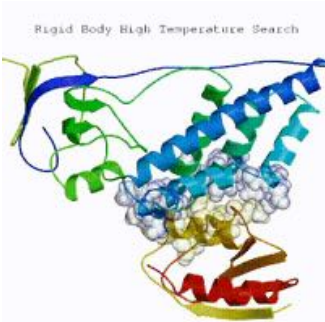


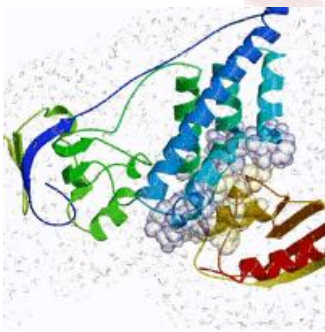
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HADDOCK


it0


it1


itw



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HADDOCK

Final score (in explicit solvent)

$$HS = 0.2 E_{elec} + 1.0 E_{vdw} + 1.0 E_{desolv} + 0.1 E_{AIR}$$

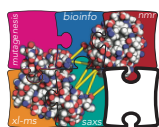


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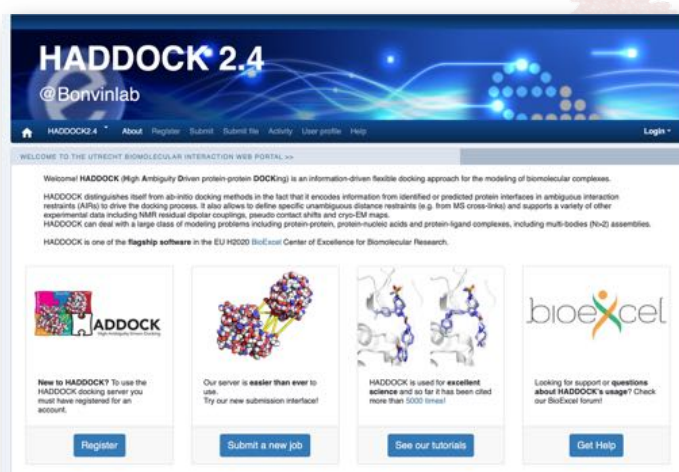
Haddock web portal



HADDOCK
High-Ambiguity Driven Docking

- > 21500 registered users
- > 330000 served runs since June 2008
- > 55% on the GRID

De Vries et al. Nature Prot. 2010
Van Zundert et al. J.Mol.Biol. 2016



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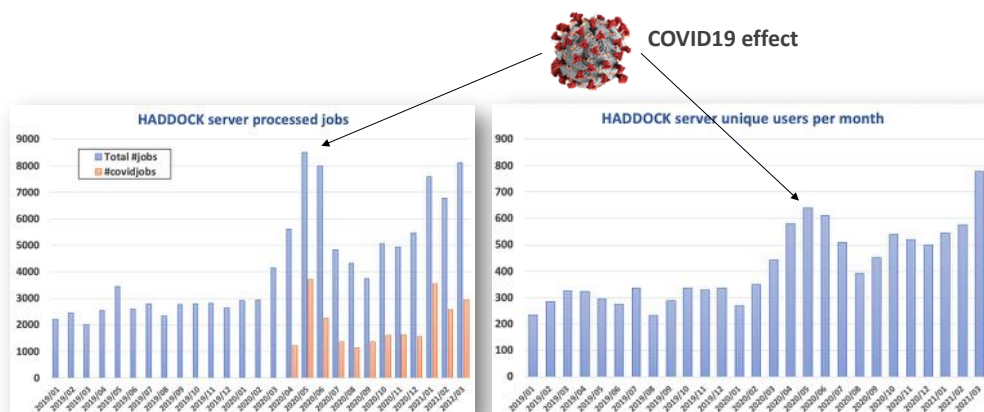
<https://wenmr.science.uu.nl>

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HADDOCK: Meeting the increased demand

The HADDOCK workflow machinery was modified to improve its efficiency and meet the increased demand (allows to run more processes in // - relevant toward exascale).



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1110001001010101010101
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Iron Piracy: NMR-based modelling of the FusA-ferredoxin complex



ARTICLE

Received 21 Jan 2016 | Accepted 21 Sep 2016 | Published 31 Oct 2016

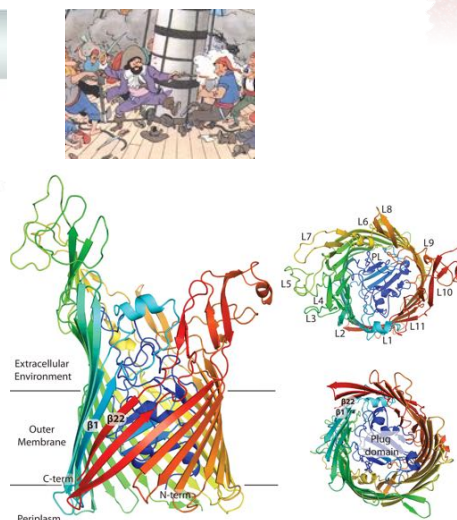
DOI: 10.1038/ncomms12304

OPEN

Structure of the bacterial plant-ferredoxin receptor FusA

Rhys Grinter^{1,2,3}, Inokentij Josts¹, Khedija Mosbahi¹, Aleksander W. Roszak⁴, Richard J. Cogdell⁴, Alexandre M.J.J. Bonvin⁵, Joel J. Milner⁶, Sharon M. Kelly⁶, Olwyn Byron⁶, Brian O. Smith⁶ & Daniel Walker¹

- **Iron import machinery in gram-negative bacteria** (pectobacterium, plant pathogen)
- **First complete crystal structure of such a receptor**



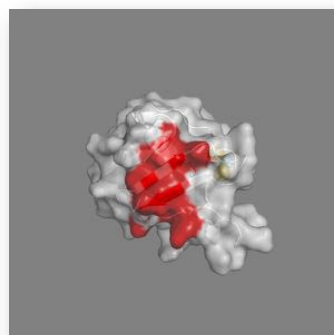
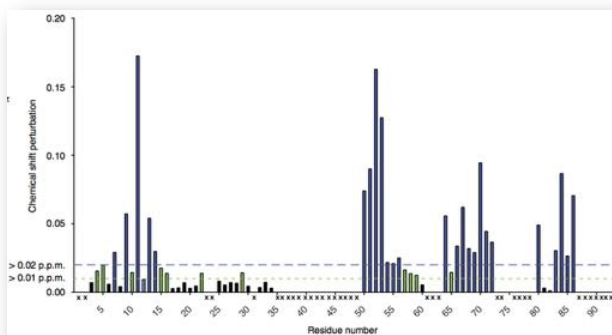
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Docking strategy

- **NMR chemical shift perturbation experiments define the binding site on ferredoxin** (which carries an iron-sulfur cluster)
→ active residues in HADDOCK



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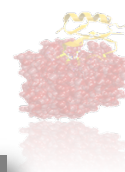
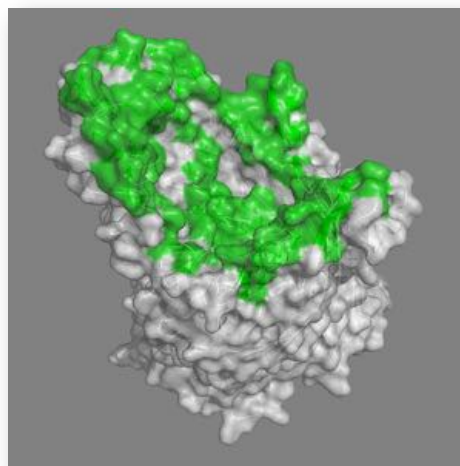
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Docking strategy

- No info for FusA (expect that the binding occurs in the extracellular part)

→ extra cellular loops defined as **passive** (which does not generate an energetic penalty if not contacted)

→ Definition of passive refined in a second docking run



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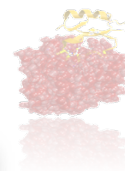
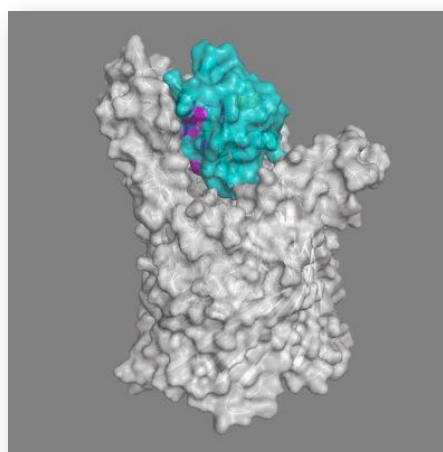
Model of the FusA-ferredoxin complex

CLUSTER 1

HADDOCK score	-137.8 +/- 2.1
Cluster size	151
RMSD from the overall lowest-energy structure	5.8 +/- 0.1
Van der Waals energy	-72.5 +/- 10.5
Electrostatic energy	-476.2 +/- 66.5
Desolvation energy	28.9 +/- 10.0
Restraints violation energy	11.2 +/- 9.62
Buried Surface Area	2524.8 +/- 175.9
Z-Score	-1.3

CLUSTER 4

HADDOCK score	-130.8 +/- 20.3
Cluster size	7
RMSD from the overall lowest-energy structure	1.4 +/- 0.8
Van der Waals energy	-70.4 +/- 17.9
Electrostatic energy	-494.9 +/- 39.9
Desolvation energy	33.9 +/- 14.7
Restraints violation energy	47.2 +/- 29.71
Buried Surface Area	2728.9 +/- 345.5
Z-Score	-1.0

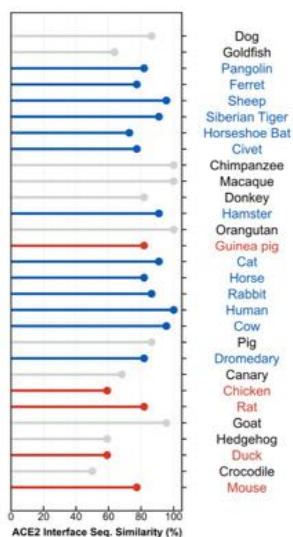


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The power of 3D structure for understanding biomolecular interactions



PLOS COMPUTATIONAL BIOLOGY

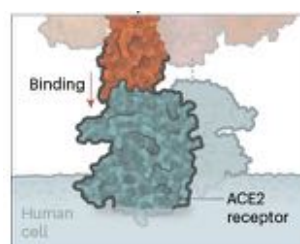
OPEN ACCESS PEER REVIEWED

RESEARCH ARTICLE

Insights on cross-species transmission of SARS-CoV-2 from structural modeling

João P. G. L. M. Rodrigues, Susana Barrera-Vilamai, João M. C. Teixeira, Marija Sorokina, Elizabeth Seckel, Panagiotis L. Kastritis, Michael Levitt

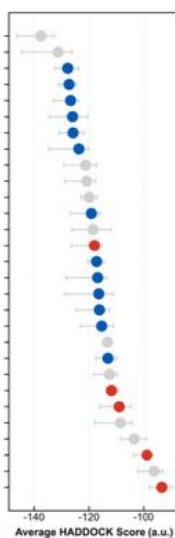
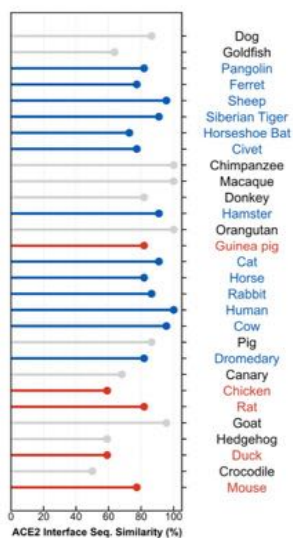
Published: December 3, 2020 • <https://doi.org/10.1371/journal.pcbi.1008449>



[Faculty of Science Chemistry]

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The power of 3D structure for understanding biomolecular interactions



PLOS COMPUTATIONAL BIOLOGY

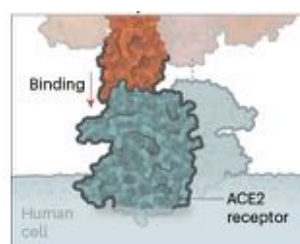
OPEN ACCESS PEER REVIEWED

RESEARCH ARTICLE

Insights on cross-species transmission of SARS-CoV-2 from structural modeling

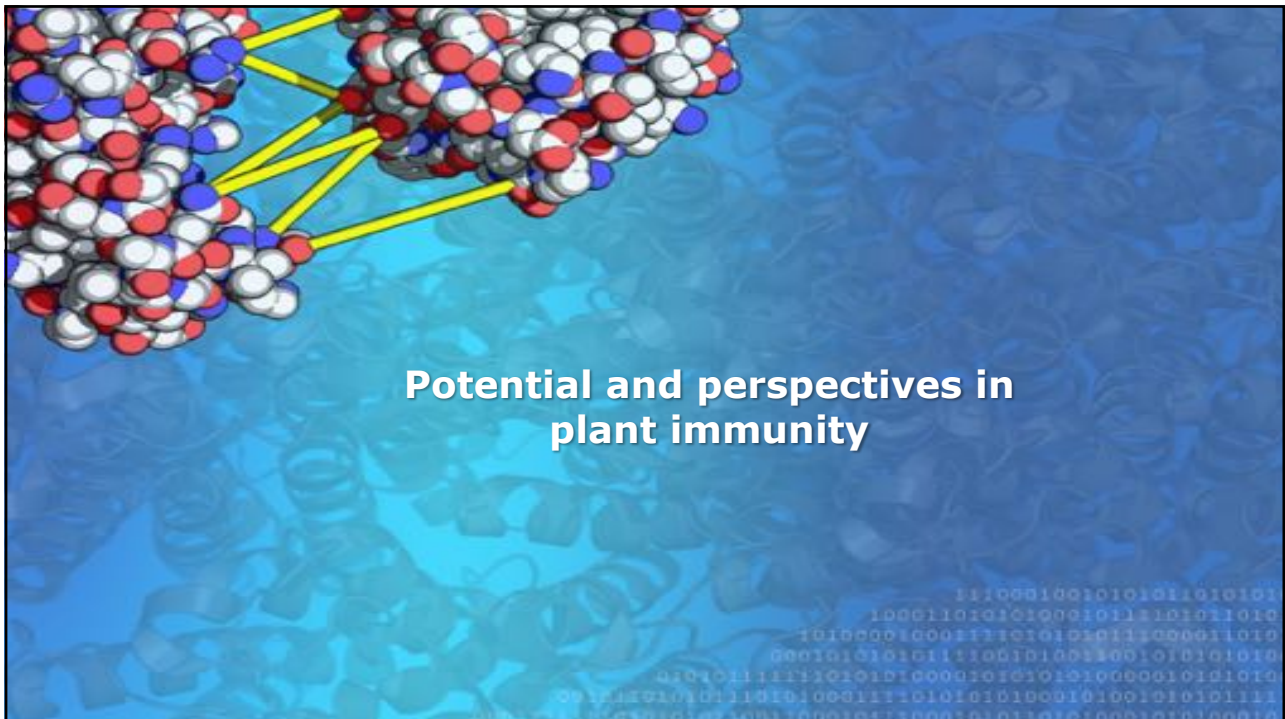
João P. G. L. M. Rodrigues, Susana Barrera-Vilamai, João M. C. Teixeira, Marija Sorokina, Elizabeth Seckel, Panagiotis L. Kastritis, Michael Levitt

Published: December 3, 2020 • <https://doi.org/10.1371/journal.pcbi.1008449>



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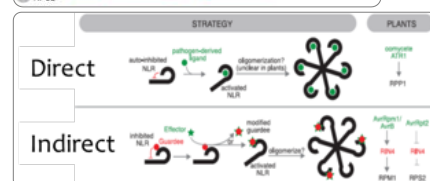
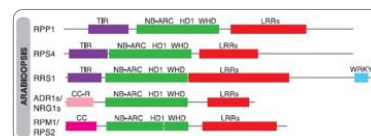


Potential and perspectives in plant immunity

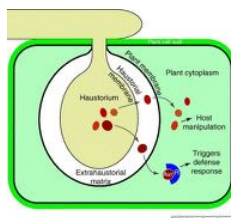
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Plant resistance (R) genes – the basics

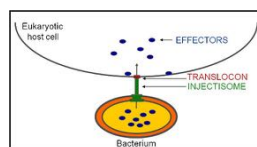
- Most code for intracellular **nucleotide-binding domain (NBD)** and **leucine-rich repeat (LRR)** proteins (**NLRs**)
- **NLRs recognize effector proteins** derived from plant pathogens
- A **recognized effector protein** is also called an **avirulence (Avr)** protein



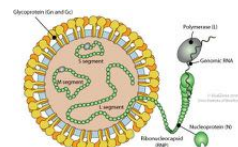
Jones et al. (2016) Science, 354



e.g. *Phytophthora spec*



e.g. *Ralstonia solanacearum*



e.g. *Tospo viruses*



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Slide courtesy of Roeland van Ham

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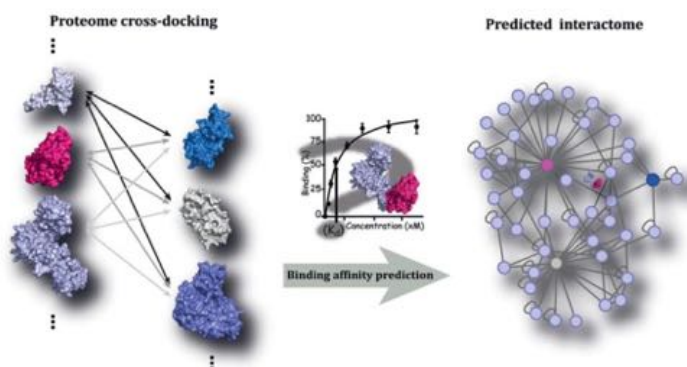
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Potential and perspectives in plant immunity

- Predict complexes between R proteins (e.g. LRR domains) and pathogen components (from sequence and/or structure)



Nature | Vol 588 | 10 December 2020 | 203



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Potential and perspectives in plant immunity

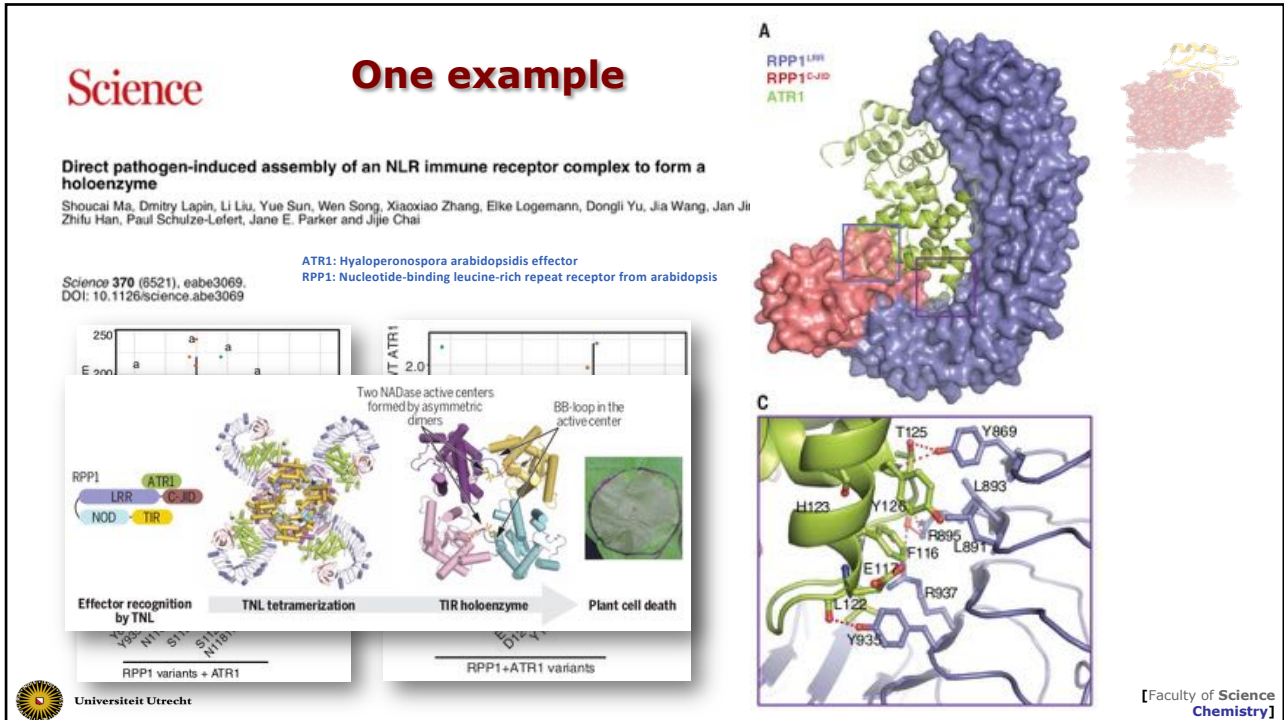
- Predict complexes between immunity proteins (e.g. LRR domains) and pathogen components
- Given the knowledge of pairs of protein interacting, identify key residues for the interaction (from sequence and/or structure)



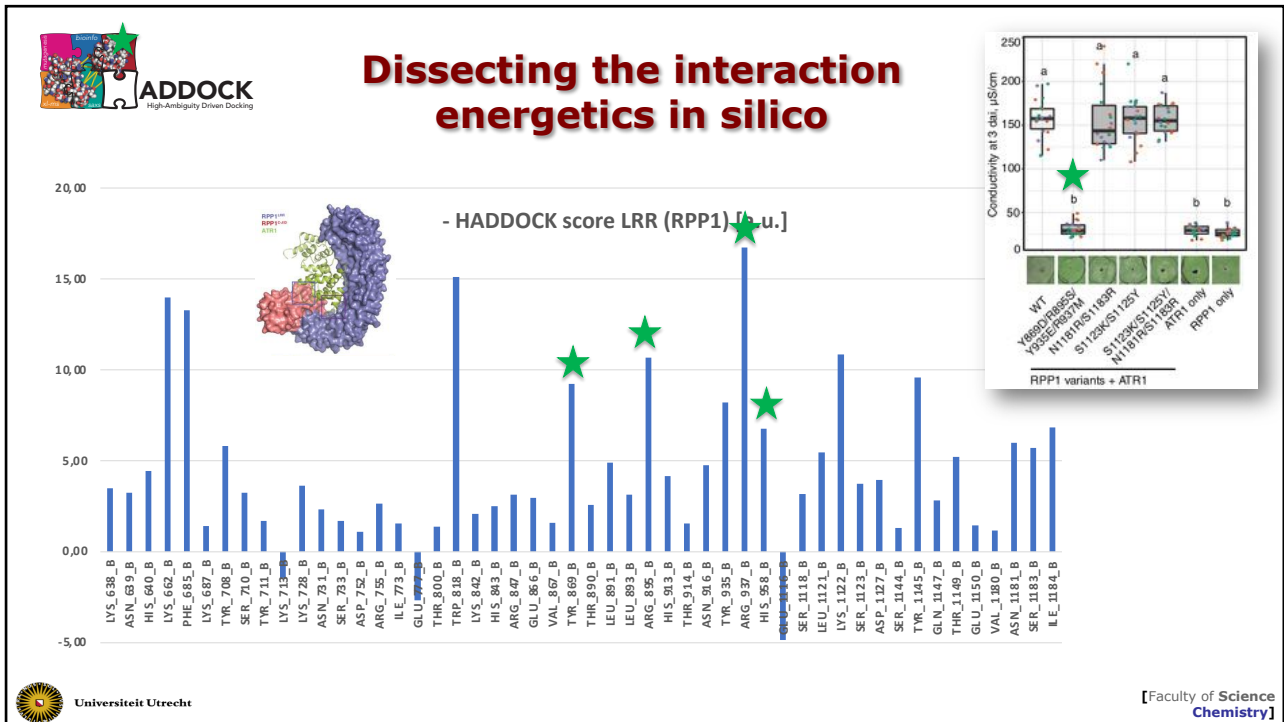
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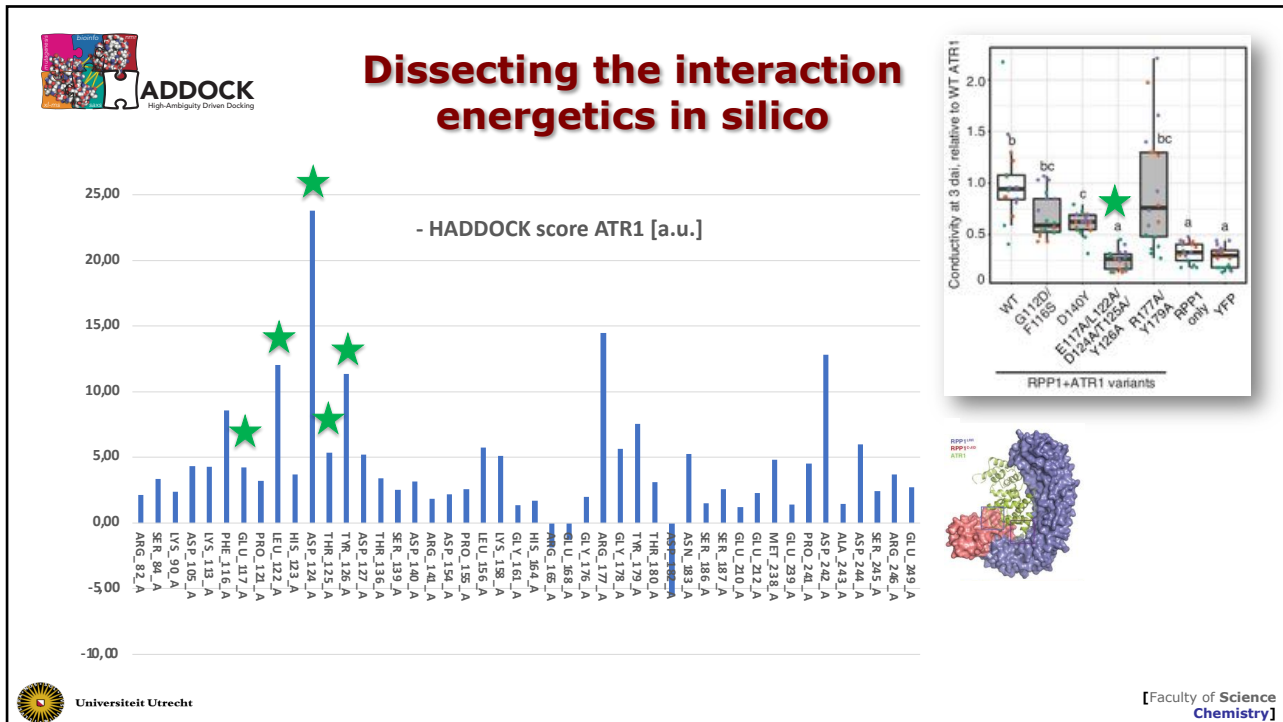
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Potential and perspectives in plant immunity

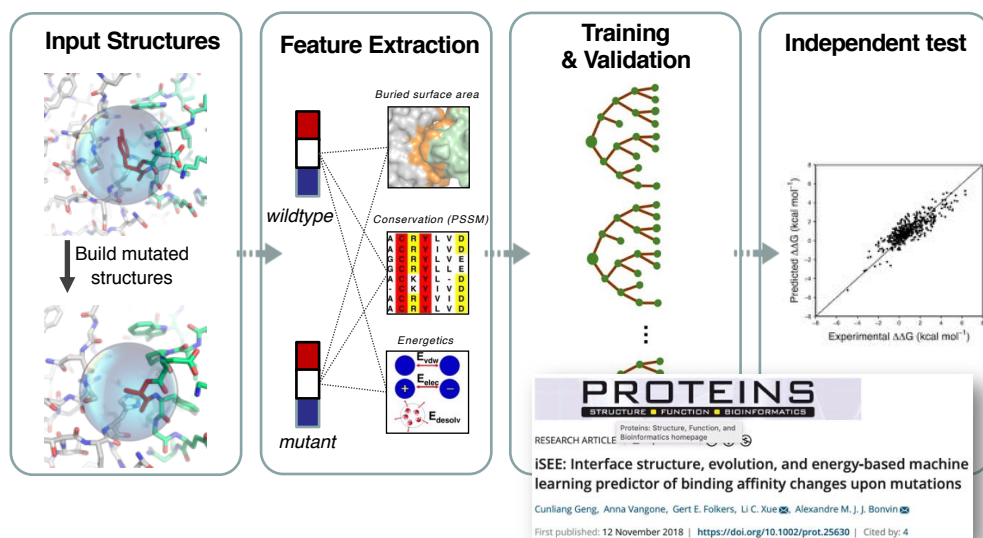
- Predict complexes between immunity proteins (e.g. LRR domains) and pathogen components
- Given the knowledge of pairs of protein interacting, identify key residues for the interaction (from sequence and/or structure)
- Given a mutation in a pathogen protein, can we select/engineer LRR domains that would restore binding (and thus induce immune response)?

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From 3D structure to prediction of the impact of mutations

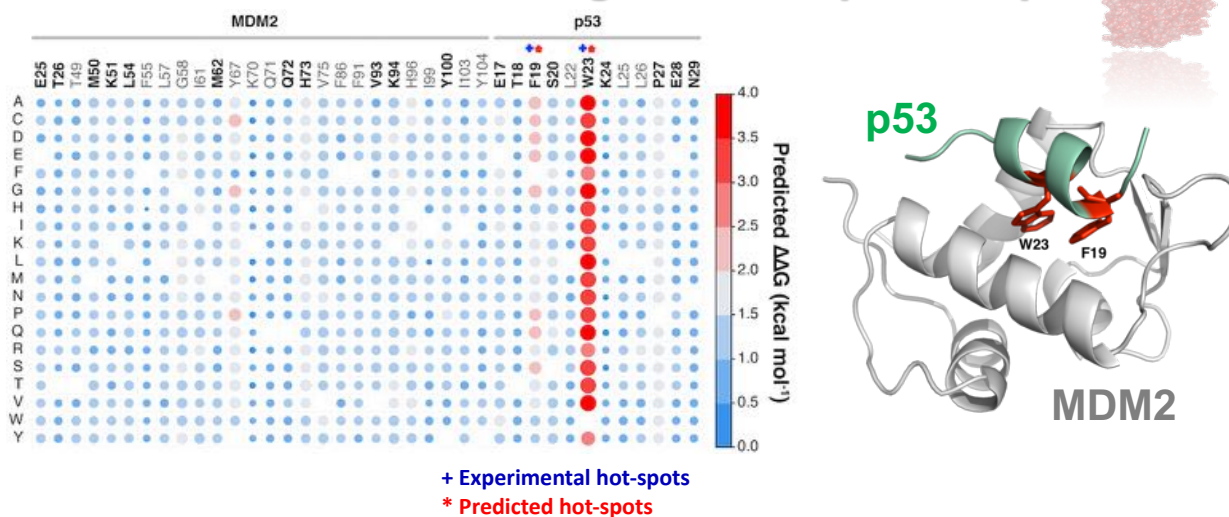


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Full mutation scanning on MDM2-p53 complex



iSEE predicted two hot-spots, which were validated by experimental data.



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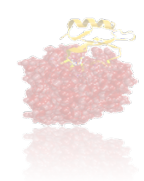
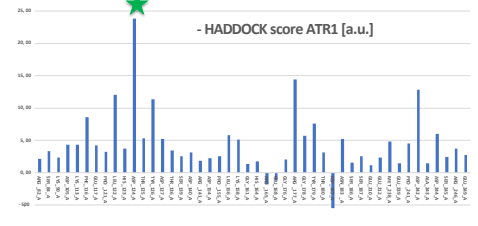
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Science **Going back to plants...**

Direct pathogen-induced assembly of an NLR immune receptor complex to form a holoenzyme
 Shoucai Ma, Dmitry Lapin, Li Liu, Yue Sun, Wen Song, Xiaoxiao Zhang, Elke Logemann, Dongli Yu, Jia Wang, Jan Jirschitzka, Zhiyu Han, Paul Schulze-Lefert, Jane E. Parker and Jijie Chai

Science 370 (6521), eabe3069.
 DOI: 10.1126/science.abe3069

As “pathogen” mutating ASP124 might help me evade recognition...

WT HADDOCK score:	-233.5 [a.u.]	Δ HS = 0
ATR1 _{D124S} HADDOCK score:	-221.6 [a.u.]	Δ HS = 11.9
RPP1 _{R895D} ATR1 _{D124S} HADDOCK score:	-228.4 [a.u.]	Δ HS = 5.1

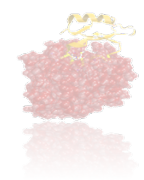
ADDOCK
 High-Ambiguity Driven Docking

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Potential and perspectives in plant immunity

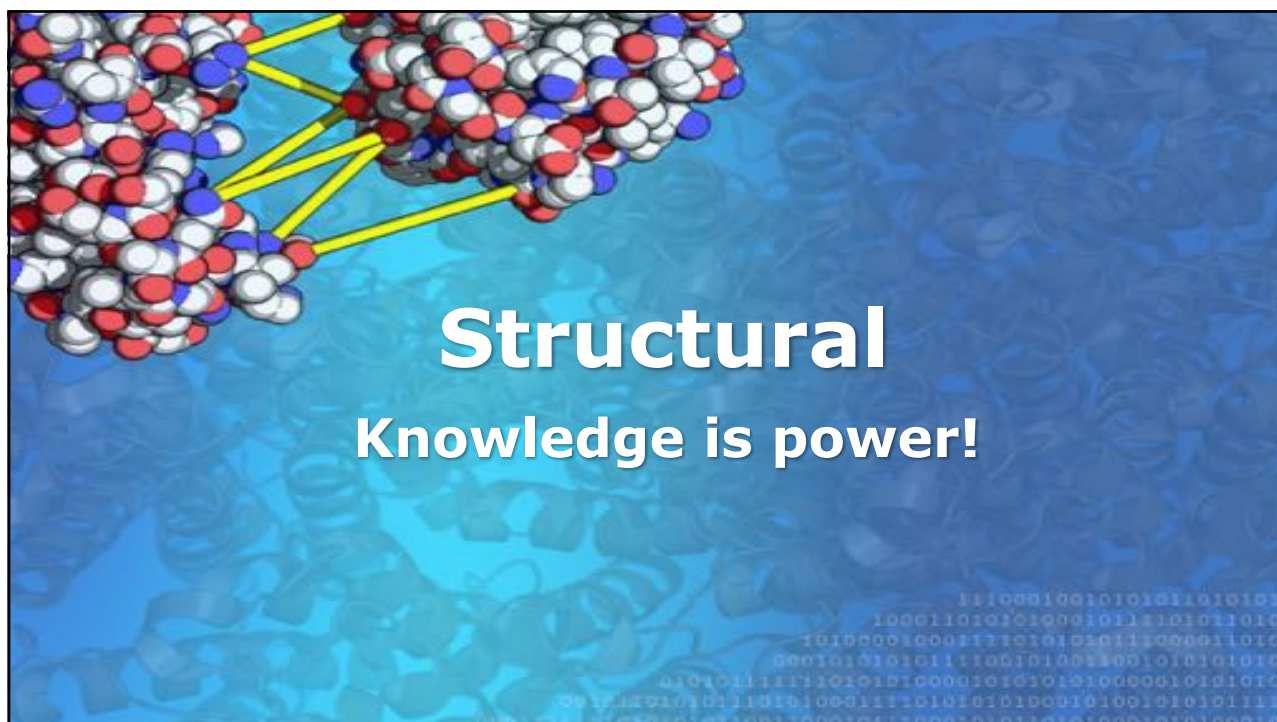


- Predict complexes between immunity proteins (e.g. LRR domains) and pathogen components
- Given the knowledge of pairs of protein interacting, identify key residues for the interaction (from sequence and/or structure)
- Given a mutation in a pathogen protein, can we select/engineer LRR domains that would restore binding?
- Both AI (deep learning) (e.g. Keygene DeepR research programme) and 3D structure can help here!

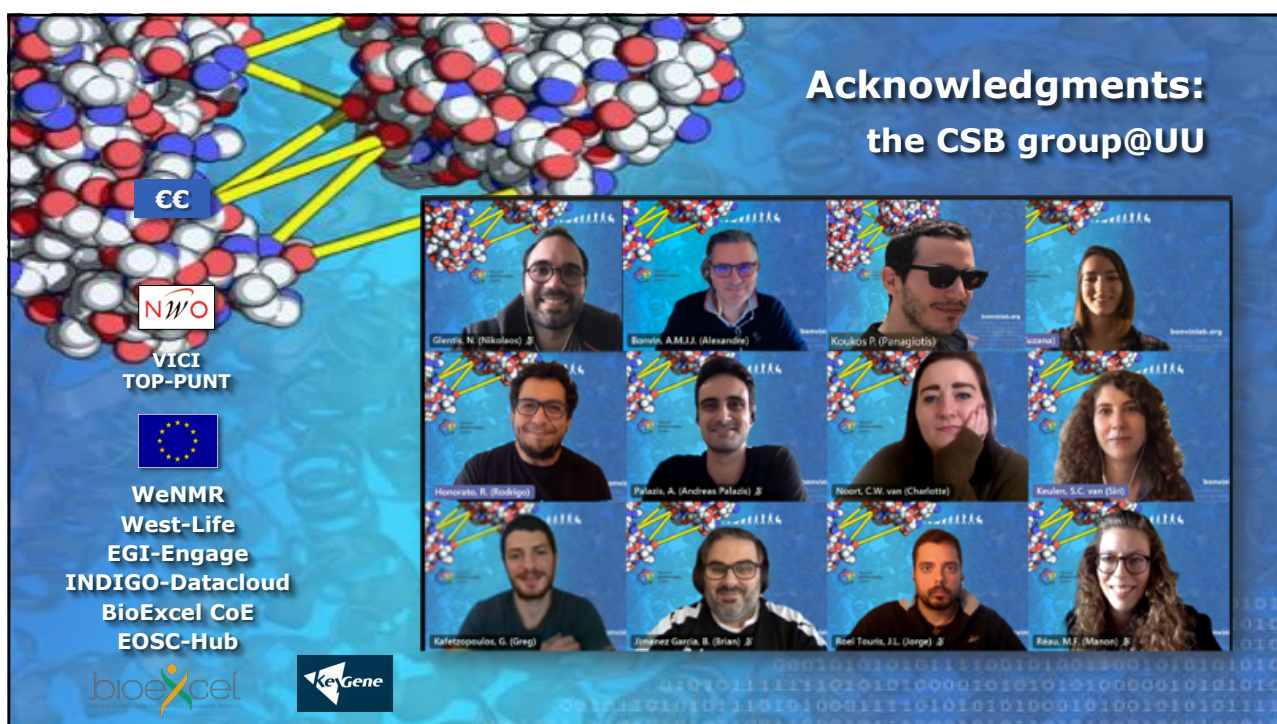
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